Development of a Clinical Trial Immunohistochemistry (IHC) Assay Using a Novel Antibody to CD38

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Introduction

CD38 is a type II transmembrane glycoprotein expressed normally in lymphoid and myeloid cells along with some non-hematopoietic tissue. CD38 is also expressed in many hematologic malignancies. A number of CD38 antibody therapies have been developed, or are under development, that induce cytotoxicity against CD38expressing cells through multiple pathways. Three drug candidates are currently in clinical testing (daratumumab, isatuximab, and MOR202) while a number of others are in preclinical development. Daratumumab is the only human monoclonal CD38 antibody that is FDA-approved for its anti-tumor activity (multiple myeloma, previously treated). Recent studies have also indicated an immunomodulatory role of anti-CD38 therapies leading to combination immunotherapy trials for various solid tumors (Van de Donk NWCJ).

With the increasing utility of CD38 as a therapeutic target, the potential for CD38 as a predictive biomarker has emerged, thus creating the need for a reliable diagnostic assay. The CD38 IHC DAK-CD38 prototype assay uses the Autostainer Link 48 automated staining platform with integrated DakoLink software. Epitope mapping revealed that the novel DAK-CD38 antibody recognizes a linear epitope. This antibody has the potential to be adapted to Dako Omnis staining platform and additional technologies such as flow cytometry.

Methods

This IHC assay is based on EnVision FLEX visualization technology using CD38 primary antibody, clone DAK-CD38, that has been developed and manufactured by Agilent Technologies. The assay staining protocol was developed for Dako PT Link and Autostainer Link 48 automated IHC platform (Figure 1). Following incubation with the CD38 antibody or the negative control reagent, specimens are incubated with a ready-to-use visualization reagent consisting of secondary antibody molecules and horseradish peroxidase molecules coupled to a dextran polymer backbone. The enzymatic conversion of the subsequently added diaminobenzidine chromogen results in precipitation of a visible reaction product localized to the antigen. Stained slides are then interpreted using a light microscope.

Development of the CD38 IHC DAK-CD38 assay was performed on three non-Hodgkin lymphoma sub-types: diffuse large B-cell lymphoma (DLBCL), follicular

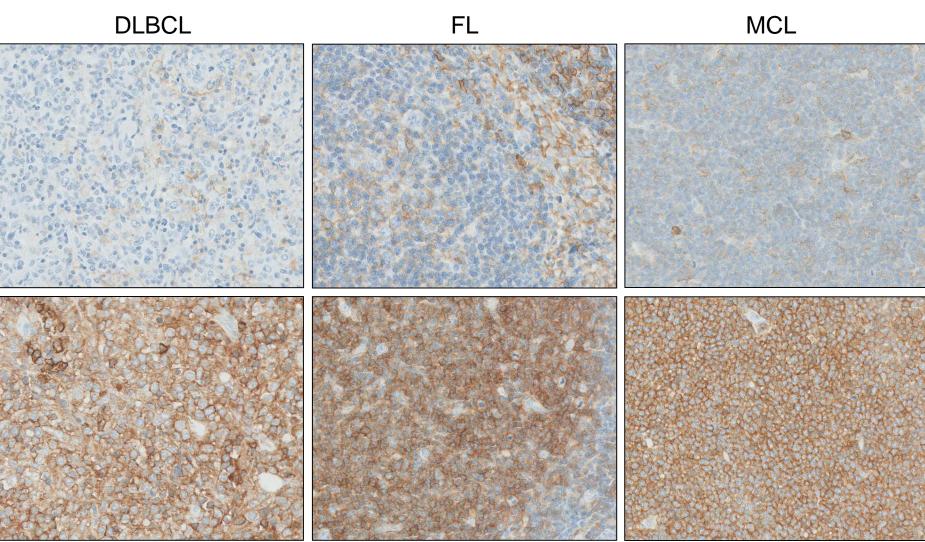
lymphoma (FL), and mantle cell lymphoma (MCL). For determination of CD38 positive staining, partial and/or complete linear circumferential membrane staining of neoplastic lymphocytes was included. Normal hematopoietic and necrotic cells were excluded from scoring. The total percent positive viable neoplastic lymphocytes with 1+, 2+, and 3+ intensity membrane staining was recorded.

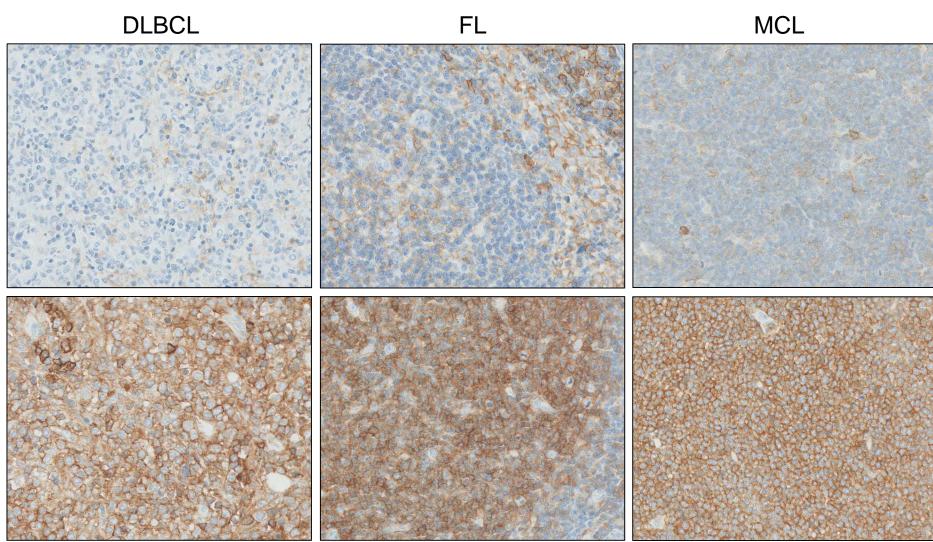


Figure 1. Autostainer Link 48 solution.

Staining:

The CD38 IHC DAK-CD38 assay provides crisp and clear plasma membrane staining with rare, minimal non-specific staining. Figure 2 shows examples of CD38 IHC DAK-CD38 assay staining in DLBCL, FL, and MCL tissues with a range of intensities (0 - 3+). Figure 3 illustrates CD38 staining of three different intensities in a DLBCL specimen.





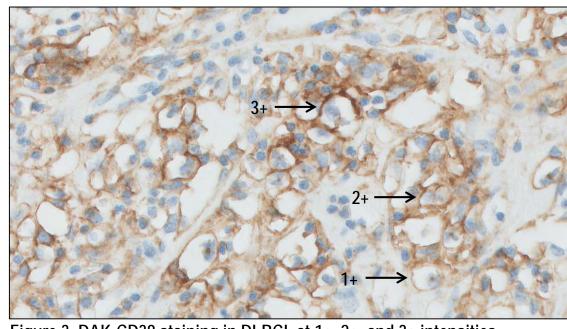


Figure 3. DAK-CD38 staining in DLBCL at 1+, 2+, and 3+ intensities.

Sensitivity:

The assay detects the CD38 protein in non-Hodgkin's lymphoma tissues and cancer cell lines.

Cell lines: 14 FFPE cell lines evaluated with the CD38 IHC assay resulted in expected staining from scientific literature. Figure 4 shows images of four cell lines with 0, 1+, 2+, and 3+ intensity CD38 expression.

Results

Figure 2. DAK-CD38 staining in DLBCL, FL, and MCL.

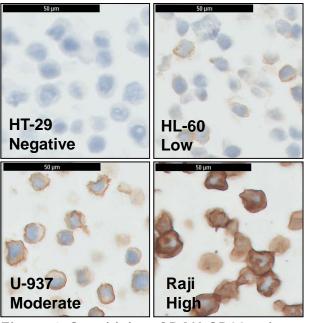
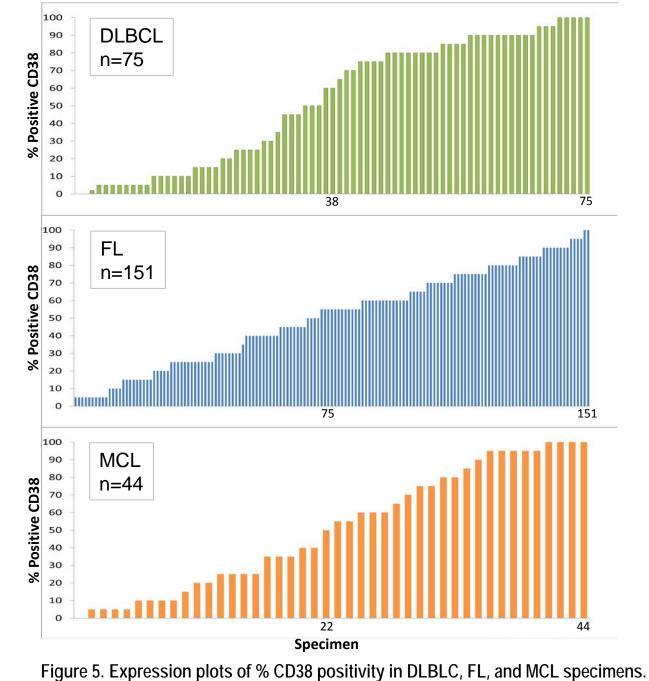


Figure 4. Sensitivity of DAK-CD38 using cell lines.

Tumor tissue: A full range of CD38 expression from 0 % to 100 % positive was observed in 75 DLBCL, 151 FL, and 44 MCL FFPE specimens (Figure 5).



Specificity: The assay demonstrates analytical specificity to CD38 using FFPE specimens other than DLBCL, FL, and MCL.

Tumor tissue: 114 different tumor tissues evaluated with the CD38 IHC assay resulted in expected staining from scientific literature. 11 tumor tissues expressed CD38 (Table 1) and 103 tumor tissues did not express CD38 (not shown).

Table 1. Specificity of DAK-CD38 using various tumor tissues.

Tissue Type	Tumor Type	# expressing CD38 / total tested		
Brain/cerebrum	Astrocytoma	4/4		
Gallbladder	Adenocarcinoma	1/6		
Head & Neck Tongue	Squamous cell carcinoma	1/2		
Liver	Hepatocellular carcinoma	1/5		
Lung	Adenocarcinoma	1/5		
Lymph node	Anaplastic large cell lymphoma	1/1		
Ovary	Serous adenocarcinoma	1/1		
Prostate	Adenocarcinoma	4/5		
Testis	Seminoma	2/2		
Uterus	Adenocarcinoma	1/3		
Abdominal Wall	Clear cell sarcoma	1/1		

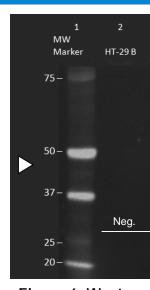
Normal human tissue: 30 different normal tissues evaluated with the CD38 IHC assay resulted in expected staining from scientific literature (data not shown).

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Results Continued

Results Continue

Western blot: cell lysates with known CD38 expression along with cells transfected with CD38 overexpressing vector (OEL), or an empty vector control, were analyzed by Western blot. A ~45 kDa band for CD38 was observed for cells known to express CD38 and was absent for cells known not to express CD38.



15 µg of protein loaded per lane.

Precision: The assay produces consistent results in normal day-to-day testing. For inter-observer precision, average positive (APA) and average negative (ANA) agreement was calculated based on a \geq 50 % CD38 expression cut-off (Table 2).

Table 2. Inter-observer agreement results with the 95 % confidence intervals in parentheses.

Agreement	DLBCL (n= 30)	FL (n= 30)	MCL (n= 30)
ANA	93.5 % (86.0-100 %)	95.5 % (89.4-100 %)	95.1 % (88.6-100 %)
APA	93.3 % (85.4-100 %)	95.7 % (88.4-100 %)	95.9 % (89.1-100 %)

Repeatability and within laboratory precision tested intra-run, inter-day, interinstrument, inter-operator, and inter-lot variables and resulted in acceptable staining variability.

Robustness: The assay produces consistent results across small variations in key assay parameters.

Studies included Target Retrieval Solution (TRS) Temperature, TRS Time, TRS pH, TRS Re-use, Post-TRS Rinse Time, Reagent Volume, Chromogen Volume, Tissue Thickness, Water Source, Run Size, Overnight Run. All robustness tests resulted in acceptable staining variability.

Conclusion

The CD38 IHC DAK-CD38 prototype assay has demonstrated acceptable sensitivity, specificity, precision, and robustness for use in detecting CD38 in DLBCL, FL, and MCL tissues.

References & Disclaimers

Van de Donk NWCJ, Janmaat ML, Mutis T, et al. Monoclonal antibodies targeting CD38 in hematological malignancies and beyond. Immunological Reviews. 2016;270(1):95-112. doi:10.1111/imr.12389.

References pertaining to expected CD38 expression are available on the digital copy of this poster via QR code download.

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3	4 Empty	5	6	7	8	9	10
P815	Vector	CD38 OEL	Daudi	U937	Raii	HL-60	Jurkat
Neg.	Neg.	High	High	Med.	High	Low	High -med.
Expected Expression							

Figure 6. Western Blot of cell lysates using DAK-CD38 antibody.

Additional References

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- 1;17(13):4232-4244.
- 20;439(3):291-296.

- 2. International Immunology. Aug 8;20(11):1369–1379.

- Apr;69(4):605-612.

- Jul 1;159(1):193-205.

1. Andorsky DJ, Yamada RE, Said J, Pinkus GS, Betting DJ, Timmerman JM. 2011. Programmed death ligand 1 is expressed by non-Hodgkin lymphomas and inhibits the activity of tumor-associated T cells. Clinical Cancer Research. July

2. Bacher I, Zidar A, Kratzel M, Hohenegger M. 2004. Channelling of substrate promiscuity of the skeletal-muscle ADP-ribosyl cyclase isoform. Biochem J. Apr 15;381:147-154. 3. da Silva CP, Schweitzer K, Heyer P, Malavasi F, Mayr GW, Guse AH. 1998. Ectocellular CD38-catalyzed synthesis and intracellular CD38-catalyzed synthesy

4. Deaglio S, Morra M, Mallone R, Ausiello CM, Prager E, Garbarino G, Dianzani U, Stockinger H, Malavasi F. 1998. Human CD38 (ADP-Ribosyl Cyclase) is a counter-receptor of CD31, an Ig superfamily member. J Immunol. Jan 1;160:395-402. 5. Deaglio S, Vaisitti T, Billington R, Bergui L, Omede P, Genazzani A, Malavasi F. 2007. CD38/CD19: a lipid raft-dependent signaling complex in human B cells. Blood. Jun 15;109(12): 5390-5398. 6. Deckert J, Wetzel M, Bartle LM, Skaletskaya A, Goldmacher VS, Vallee F, Zhou-Liu Q, Ferrari P, Pouzieux S, Lahoute C, Dumontet C, Plesa A, Chiron M, Lejeune P, Chittenden T, Park PU, Blanc V. 2014. SAR650984, A novel humanized CD38targeting antibody, demonstrates potent antitumor activity in models of multiple myeloma and other CD38⁺ hematologic malignancies. Clin Cancer Res. Sep 1;20(17):4574-4583. 7. Dussault N, Ducas E, Racine C, Jacques A, Pare I, Cote S, Neron S. 2008. Immunoglobulins involves increased phosphorylation of extracellular signal-regulated kinases 1 and

8. Ellmark P, Belov L, Huang P, Lee CS, Solomon MJ, Morgan DK, Christopherson RI. 2006. Multiplex detection of surface molecules on colorectal cancers. Proteomics. Feb 16;6:1791-1802. 9. Gabay C, Ben-Bassat H, Schlesinger M, Laskov R. 1999. Somatic mutations in the rearranged V_k genes of B-non-Hodgkin's lymphoma cell lines. Eur J Haematol. Sep;63(3):180-191. 10.Ghaderi AA, Amirghofran Z. 1994. CD38 Molecule-A multilineage glycoprotein and its unique expression on plasma cells. Medical Journal of the Islamic Republic of Iran. Aug;8(2):109-113. 11.GolayJ, Erba E, Bernasconi S, Peri G, Introna M, The A-myb gene is preferentially expressed in tonsillar CD38+, CD39-, and slgM- B lymphocytes and in Burkitt's lymphoma cell lines. J Immunol July 15, 1994, 153 (2) 543-553. 12.Gutierrez M, Cherney B, Hussain A, Mostowski H, Tosato G, Magrath I, Bhatia K. 1999. Bax is frequently compromised in Burkitt's lymphomas with irreversible resistance to Fas-induced apoptosis. Cancer Research. Feb 1;59:696-703. 13.Hamilton MS, Ball J, Bromidge E, Lowe J, Franklin IM. 1990. Characterization of the biology of multiple myeloma. British Journal of Haematology. Mar, 8;75(3):378-384. 14.Hermann M, Scholman HJ, Marafioti T, Stein H, Schriever F. 1997. Differential expression of apoptosis, Bcl-x and c-Myc in normal and malignant lymphoid tissues. Eur J Haematol. Jul;59(1): 20-30. 15. Horenstein AL, Stockinger H, Imhof BA, Malavasi F. 1998. CD38 binding to human myeloid cells is mediated by mouse and human CD31. Biochem J. Mar 15;330(3):1129-1135. 16.Iwata K, Ogata S, Okumura K, Taguchi H. 2003. Expression of CD38 in human promyelocytic leukemia HL-60 cell line during differentiation by niacin-related compounds. Biosci Biotechnol Biochem Aug;67(8)1836-1839. 17.Klein U, Klein G, Ehlin-Henriksson B, Rajewsky K, Kuppers R. 1995. Burkitt's lymphoma is a malignancy of mature B cells expressing somatically mutated V region genes. Molecular Medicine Jul;1(5):495-505. 18.Liu Q, Kriksunov IA, Graeff R, Munshi C, Lee HC, Hao Q. 2005. Crystal structure of human CD38 extracellular domain. Structure. Sep;13(9):1331-1339. 19. Malavasi F, Deaglio S, Damle R, Cutrona G, Ferrarini M, Chiorazzi N. 2011. CD38 and chronic lymphocytic leukemia: a decade later. Blood. Sep 29;118(13):3470-3478. 20.Morra M, Zubiaur M, Terhorst C, Sancho J, Malavasi F. 1998. CD38 is functionally dependent on the TCR/CD3 complex in human T cells. FASEB J. May;12(7):581-592. 21. Munshi CB, Graeff R, Lee HC. 2002. Evidence for a causal role of CD38 expression in granulocytic differentiation of human HL-60 cells. J Biol Chem. Dec 20;277(51):49453-49458. 22.Musso T, Deaglio S, Franco L, Calosso L, Badolato R, Garbarino G, Dianzani U, Malavasi F. 2001. CD38 expression and functional activities are up-regulated by IFN-gamma on human monocytes and monocytic cell lines. J Leukoc Biol.

23.Sconocchia G, Titus JA, Mazzoni A, Visintin A, Pericle F, Hicks SW, Malavasi F, Segal DM. 1999. CD38 triggers cytotoxic responses in activated human natural killer cells. Blood. Dec 1;94(11):3864-3871. 24.Webster R, Pulford K, Jones M, Micklem K, de Waele M, Mayne K, Tse A, Mason DY. 1993. ZB51: a monoclonal antibody reactive with human plasma cells. Br J Haematol. Aug;84(4):602-607. 25.Zheng X, Ding N, Song Y, Feng L, Zhu J. Different sensitivity of germinal center B cell-like diffuse large B cell lymphoma cells towards ibrutinib treatment. Cancer Cell Int. Apr 3;14(1):32. 26.Zubiaur M1, Izquierdo M, Terhorst C, Malavasi F, Sancho J. 1997. CD38 ligation results in activated protein-70 signaling pathways in Jurkat T lymphocytes. J Immunol.

27.Van de Donk N, Monoclonal Antibodies Targeting CD38 in Hematological Malignancies and Beyond Immunological Reviews 2016; 270; 95-112 28.De Weers M, et al. Daratumumab, a Novel Therapeutic Human CD38 Human Antibody, Induces Killing of Multiple Myeloma and Other Hematological Tumors J Immunol 2011; 186:1840-1848